



The

June 2011

BUZZWORD

West Sound Beekeepers Association <http://www.westsoundbees.org/>
Proudly serving bees, their keepers, and the public in Kitsap County, WA
(Interactive Newsletter is in the works!)

A

**Special Thanks to
Mary Felts!**

**6 PM Bee-ginner's Beekeeping Class
June 7 Last Class and open book exam**

**June 21, 2011 7 PM Regular Meeting
Program:
David Mackovjak**

"inexpensive" Beekeeping

**Meeting Schedule:
Bee-ginning Beekeeping
Class**

6PM Tuesday 7 June 2011

Regular Meeting

7PM Tuesday June 21 2011

**Steering Committee
Meeting**

7 PM Tuesday 05 July 2011

**Queen Rearing Group
Meets after the Steering
Committee**

**Meetings at Stedman's
Beekeeping Supplies in
Silverdale**

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Advanced Beekeeping Course
**Advanced Beekeeping in
the Pacific Northwest 2011**
Saturday July 9, 2011
Silverdale, Washington

Details page 17

**Yes, a reminder the
WSU field day is
June 24 and 25 in
Pullman**

see page 15

WSBA Business Meeting Minutes June 7, 2011

Submitted by Christine Torres, Secretary

Called to order at 7:05 pm; All officers in attendance

New Business:

- T.J. Jorgenson will contact Puget Sound Beekeepers Assn. to see if interested members can join them in their fireweed honey harvest location(s).
- Topic of next general meeting: Inexpensive beekeeping. Led by David Mackovjak
- Kim Redmond presented \$100.00 donation to Queen rearing group and reported another \$100.00 donation will be coming from Port Ludlow Resort.
- Maple seedlings available for a donation to the queen rearing group. See Kim Redmond.
- David Mackovjak presented preliminary outline for advanced beekeeping course.

Meeting adjourned: 8:10pm

Message from the President: I'm out of town on vacation and so wsba has a president on hiatus and who was distracted in preparations for it and did not prepare a message for this month.

Apologies.

This months meeting will be on inexpensive beekeeping.

Apprentices took their exams.

I have gotten a dozen emails at least from people who starved their bees this year by ending feeding of packages far too early. Perhaps my message would only be a reminder to feed during the dearth.

But suffice to say, I'm out of the office until the end of June. -TJ



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NW Beekeeping Basics –May/June

Examine each colony every 9 days and treat as in April. Watch out for the lull in nectar production between the maples and the blackberries. Large colonies can quickly exhaust stores of honey and pollen during a week of inclement weather and cannibalize brood or starve! Without feed sources around, your bees, which look big and strong and hard working, can actually starve out in the first three weeks in May. So, watch your bees carefully and don't let colony stores get below 15 lbs. Feed syrup and pollen patties if necessary, especially bees started on foundation. Basically, feed until they stop eating (fat little bees!)

Give the single story colonies getting full of bees a second story. Make sure you give the bees another box in time to prevent swarming. Give supers with foundation, but only to those colonies that are working in the supers. As long as there is nectar coming in, the bees will draw the foundation into comb.

Place new supers directly over the queen excluder, if you are using one. Remove and extract the supers containing well ripened honey.

Order queens for July delivery. Requeening is your best management tool and losing a queen now or having a failing queen now will dramatically effect your honey crop. You will need queens to make your nucs even if you do not plan to expand the number of hives you have. Having nucs on hand will allow you to successfully requeen colonies late into the year.

Keep in mind that brood laid in late May are not going to be old enough to be field bees during the honey flow. So, have new strong queens on hand to make your nucs.

Check your stored comb for possible wax moth infestation – like rust, wax moths never seem to sleep.

Make your plans for your county and state fair entries.

Now is the swarm season when you hope that your neighbor beekeeper is not as good a beekeeper as you are and you can catch swarms from their hives while your good management practice keeps your hives from swarming. Swarm control is probably one of the least successful areas of beekeeping. Swarming is the natural way for bees to ensure the survival of their species, and like all teenagers, the instinct to reproduce is very difficult to control. One of the best ways to minimize swarming is by requeening your hives. A swarm is a portion of your bees leaving with the old queen, and this tends to be with queens in their second year. So by requeening with a new young queen, you not only assure yourself of a strong queen, you cut down on the chance of her swarming.

A second technique is to reverse your hive boxes. The bees will start the swarm process when the queen starts to run out of perceived space in which to lay. Since she tends to only move upward, she does not use the space available below the brood cluster. By reversing the boxes, you force the bees to reorganize their stores and therefore create new space for the queen to lay in. Another method is to remove queen cells as they appear. But this rarely works since it's next to impossible to find all the queen cells and usually the bees have committed to swarming no matter what and just build more cells. Additionally the constant intrusion into the hive will stress the colony greatly. The simple act of adding supers can also make the colony feel they have plenty of room. Give supers with foundation, but only to those colonies that are working in the supers. Place directly over the queen excluder, if you are using one.

With swarming in mind, examine your colonies about every 9 days(on the 10th day a queen could emerge from a larva jumpstarted right about the time of your last visit. Then it's Sayonara! Adios! Farewell! Sometimes the bees leave before the queen emerges!) To inspect: Set off the supers; tilt up the second story, give a puff of smoke, and look for queen cells along the bottom of the brood comb. If you find only eggs or larvae in the queen cells and the hive is crowded with bees, remove all the cells -- top and bottom. Put the hive body containing mostly worker bees or larvae on the bottom board, and the other containing mostly sealed brood on top. Next put on the queen excluder and add supers to provide 10 to 15 empty full depth comb, or their equivalent in the supers. Place the empty comb directly on the excluder.

If you find sealed or ripe queen cells, or possibly hatched ones, divide the colony. Set the top body, usually with most of queen cells, onto a bottom board and cover with a lid. Put this hive on a separate stand. Remove all queen cells from the lower body; put on the excluder and add supers to provide 10 to 15 empty full depth combs, or their equivalent, above the excluder. Several hours later, look for the queen in the divide. If you find a virgin queen (no eggs laid), let her remain, but if you find the old queen pick her up and let her run into the entrance of the colony on the old stand. Let the queen cells hatch in the divide. If you find no attempt to rear queens, and the hive is full of bees, examine the supers and add more to provide 10 to 15 empty full depth combs, or their equivalent, above the excluder.

Think about adding a screen bottom for better ventilation, if you're not already using one, for better ventilation and mite control that doesn't cost any money after installation. Some beekeepers use a additional 3/4-1" hole in the upper box as a entrance and to provide additional ventilation. Bees seem to love these!

Keep on the lookout for American Foulbrood (AFB). You can spot it by piercing sunken capped brood with a tooth pick (or similar tool) and looking for "stringy" filling in place of brood. A second method is to hold the comb by the top bar at an almost flat angle, with the sun to your back, look for dark or black scale on the bottom of cells in the brood area. AFB can be treated different ways. If using antibiotics, wait until honey supers are off to avoid contaminating honey. The comb may have AFB spores, that is something to think about! If it's a very small outbreak it may be possible to remove the affected frames and treat with Terramycin. If treating with Terramycin do not treat with supers on that you intend to extract for honey. Some strains of AFB are terramycin-resistant and the stronger and more residual antibiotic Tylosin can be used. In the event of a full-blown outbreak the recommended traditional action is to destroy the colony and burn the equipment. Non-traditional methods of combating AFB without antibiotics include "Shook Swarming" where all the bees are shaken into clean equipment without drawn comb, just foundation. Equipment except for the comb itself (which should still be destroyed) can be reclaimed by sterilization using an autoclave, gamma irradiation, or a hot paraffin dip. Also baking, scorching, repainting and bleach are helpful techniques for reclaiming equipment.

If you have been fortunate and the honey flow has been good, add new supers under supers with capped honey. Remove and extract the supers containing well-ripened honey.

May 2011 Treasurer's Report

In the month of May 2011 we added one new member and one more member paid their annual dues. We funded the balance of Nicole Froyd's 2010 scholarship by issuing a reimbursement check based on her purchase of a package of bees. Other expenses included stamps, newsletter software and honey sticks to hand out at Ecofest.

Income for May was:

Beginner Study Guides ~\$10
2011 Dues ~ \$48
Donation ~ \$1
Interest ~ \$.55

TOTAL INCOME: \$59.55

Expenses for May were:

2010 Scholarship Balance ~ \$52.87
Newsletter expenses (software) ~ \$87.79
Stamps ~ \$47.50
Fair expenses ~ \$20.00

TOTAL EXPENSES: \$208.16

NET INCOME FOR May 2011: <\$148.61>

| | |
|--|-------------|
| Checking account balance @ 5/31/2011 ~ | \$ 2,990.34 |
| Savings account balance @ 5/31/2011 ~ | \$ 6,381.86 |
| Total Cash on Hand @ 5/31/2011 ~ | \$ 9,372.20 |
| Undeposited Funds @ 5/31/2011 ~ | \$ 0.00 |

2011 Scholarship Liability (Warren Carlson) Balance @ 5/31/11~ \$3.43

The WSBA Scholarship Fund value @ 5/31/2011 = \$258.13

DONATIONS

Directed and/or General donations that support all aspects of the Association are always welcome ~ in any amount!

Kim Redmond
Treasurer

Queen Rearing Group Update



Brand new 2011 WSBA Queen

Purpose: The Queen Rearing group is to provide an opportunity for members of WSBA a venue to learn and explore the art of queen rearing and produce quality queens.

2011 Goals for the Queen Rearing group of West Sound Beekeeping Association (WSBA):

1. Explore and document different physical methods of queen rearing
2. Educate both the Queen Rearing group and the WSBA on queen rearing
3. Provide quality queens to the Queen Rearing group and WSBA
4. Investigate and conduct hygienic testing
5. Explore drone management methods
6. Invite at least two other local beekeeping associations to our meetings
7. Artificially inseminate a queen (stretch goal)

The Queen Rearing group meets on the first Tuesday of the month following the WSBA business meeting held at Stedmans. The business meeting starts at 7 PM and the Queen Rearing meeting will start at approx 8 pm or after the business meeting ends.

Part 1 – Tues, 6/7/11 (2000 to 2200) Class on Instrumental Insemination



Following the normal West Sound Beekeepers Association business meeting, the Queen Rearing Group met. There were approximately 20 in attendance and they were interested in the training given by George Purkett on Instrumental Insemination. George presented a slide show of the April 20-21 advanced workshop on queen insemination course from the UC David taught by Susan Cobey. The course was a practical hands-on approach providing detailed instruction and hands-on practice of semen collection and insemination of queens. Various types of insemination instruments

are displayed and reviewed. The preparation and care of virgin queens and drones was presented. Basic bee genetics and various breeding systems were reviewed. George had a great slide show and answered lots of great questions that evening. We are hoping to get the club's Instrumental Insemination equipment within the next 30 days.

Part 2 – Thurs, 6/10/11 (1800-1900) Cloake Board setup

On Thursday afternoon we gathered at the apiary and setup one of the hives for the use of Cloake Board. The Cloake Board Method takes advantage of both a queen-less and queen-right system. If you recall from last month's training, Queen cells started in a queen-less state tend to have a higher rate of acceptance, and those reared in a queen-right state tend to produce higher quality cells. Hence, the popularity of the starter and finisher methods used. Another key component of rearing premium queen cells is a minimal amount of disturbance. Moving developing larvae between starters and finishers interrupts the critical and intensive larval feeding stage. The Cloake method eliminates the need for this practice. Experiments have shown that a developing queen larva receives 1600 feeding visits from nurse bees, compared to the 143 feeding visits received by a worker larva. Thanks to the ½ dozen or so folks who came out to watch and ask questions.



The lower super has the queen and a queen excluder



A removable divider is installed



Cloake hive all set up and ready for grafts



Banked queens from last month

Part 3 – Sun, 6/12/11 (1300-1630) Grafting session, queen cage building, and queen marking.

We met at Stedman's on Sunday, June 12, 2011 at 1 pm on a nice sunny day. This was a perfect day for our second grafting session of the season! We had a great turnout of club members to include Jean, Tom, George, David, Melissa, Charles, Suzanne and the entire (4 members) Carlson family. The first order of business was to briefly review the processes of grafting. We explained and answered questions about the equipment, techniques, conditions, process, etc. It is all about practice, practice, practice.....

| Name | Total No. | Good graft | % |
|------------------|-----------|------------|-----------|
| Jean Ball | 13 | 9 | 69 |
| Jim Carlson | 12 | 10 | 83 |
| James Clements | 11 | 9 | 82 |
| Morgan Carlson | 15 | 11 | 73 |
| Roseanne Carlson | 14 | 10 | 71 |
| Selena Clements | 17 | 9 | 53 |
| Total | 82 | 58 | 71 |

Last month's grafting results are in. We did much better than was expected for our first graft session. I was expecting about a 40% success rate and overall we average over 70%. Wow!! Everyone should be proud of their grafting ability after just one training class. (We must have some really good instructors©)The award of "Master Grafter" goes to Jim Carlson who grafted an outstanding 83% success rate!! Congrats and well done!!

Like last month, everyone was given the opportunity to try their hand at grafting the newly hatched larva and placing them into the queen starter cups. We had several different grafting tools on hand and everyone was encouraged to try all the different models. The idea was to locate a newly hatched larva, ideally less than 24 hrs old. They were really small so we were using a light assisted magnifying glass. It was important to keep the larva in the same orientation and also from drying out so we had moist towels over the brood comb. Everyone got a chance to try grafting using both the Chinese and German tools.



George demonstrating the basics in grafting (< 36 hrs old)



Charles looking for the correct larva (< 36 hrs old)



Suzanne experimenting with different grafting tools



Jim preparing a row of grafts

Once again George fired up his wax queen cell cup maker. It consisted of a hot pot converted into a double boiler in which he had melted bees wax. Then he had dowels that had their ends shaped of which he would first dip in cool water then repeatedly dip in the beeswax four or five times and then quickly melt them to the cell board.



George demonstrating how to make wax queen cups



Melissa making a row of wax queen cups

We also gave everyone the opportunity to make queen cages. We will use these to place the queen cells into a day or so before the queen emerges. This prevents them from fighting each other and allows us to bank them until we are ready to use them. George brought some wire mesh, tin snips, a dowel to form the mesh around and a soldering gun. Once a couple were made it was pretty easy to mass produce them.



George soldering a queen cage



Jean building her first queen cage

We brought in a ½ dozen or so queens so the members could practice the technique of marking queens. Marking a queen bee will allow the beekeeper to locate her easily by the now distinguishing spot on her back. Specialty markers can be purchased from a beekeeping supplier, but similar ink can be found more easily at a local arts and crafts store. Just make sure to let the ink dry before releasing her, otherwise her colony will clean it right off.

There is an International Color Code system for marking queens. Do not just mark your queens any ole color. You'll forget what year you started with them. Here's how the color code works: Year ending: 5 or 0 Blue, 6 or 1 White, 7 or 2 Yellow, 8 or 3 Red, 9 or 4 Green. So for 2011 we mark our queens with a white marker.



Melissa marking her first queen



Suzanne proudly showing off her marked queen



CO2 to knock out the queens for marking



Jean showing proper technique to pick up a queen

All this queen rearing training worked up an appetite and the Jim and Roseanne were kind of enough to bring pizza for everyone. We took a break swapping stories and enjoyed the sun shine. Thanks!!



Once everyone felt satisfied with all the grafting practice we went back out to the apiary and pulled several frames from the same hive (#4). We pulled the queen cups that I had put in the Cloake board hive 24 hrs the day before. We selected six final grafters, so by next week we'll see who our "Master Grafter" is for this month's queen rearing grafting.

Once again the queen rearing group grafted over 90 queen cells and didn't end up leaving until late afternoon. I think everyone once again had a great learning opportunity. A special thanks goes to George for bringing in all his equipment!!

At our next WSBA meeting, on June 21st we will need to go back out to the finishing hive and place the developed queen cells into either holding pens or nucs. We are selling the queen cells for \$4/cell or \$2/cell if you are a participating member of the queen rearing group. This is a great deal!! Remember everyone should have a nuc or two in standby in case you lose your queen.



Lecture Schedule: We outlined our Queen Rearing lecture schedule and the following lectures will be presented during our upcoming meetings.

5 Apr – Drone Management – Jim Carlson
3 May – Cloake Board and grafting – David Mackovjak
7 Jun – Instrumental Insemination – George Purkett
5 Jul -- Hygienic Testing – Paul Lundy

Hands-on-Demonstrations: A schedule of hands-on-demonstrations and skill events was also discussed and confirmed (dates subject to weather and hive strength)

Thurs, 7 July 11 – Cloaking setup demonstration, 6 pm Stedmans
Sunday, 10 July 11 – Hygienic Testing & Queen grafting demo, 1-3 pm Stedmans

Thanks and looking forward to seeing a great turnout at our next Queen Rearing meeting on 5 July when Paul will discuss how we can test a hive for its hygienic behavior. Come join all the fun!!

The Queen Rearing group is open to all members who are interested in learning about the art of queen rearing or would just like to learn more about bee keeping. To be included in queen rearing correspondence please send me your name, phone number and email address.



David Mackovjak
Queen Rearing group leader
trimack@prodigy.net
360-340-0381



Beekeeping in Silverdale, WA

Letters From The Librarian

Through the Warre' Window, Part 1.

Last week (May 24) I went out on a swarm call and retrieved a very nice swarm to populate my brand new freshly painted Warre' hive. Actually, I have two coats of primer on it and was debating on a color scheme when the call came in so my artistic side will have to wait. It is currently two boxes deep and the window is on the back of the bottom box. The frames have angled top bars, partial sides and no bottoms. The cluster did not start in the center like I would have, they are more to one side. It looks like they are going to start drawing comb in the lower frames below the frames they are currently on before expanding across the upper box. Again, this is not how I would have done it.

There is a screened bottom on the hive and I am surprised at the amount of wax flakes that fall through. I wonder if the amount of wax flakes will correlate with wax building and nectar flow later in the season. I also haven't spotted any mites under the screen yet.

The other thing to watch is their comings and goings. When they leave, they fly down to the bottom and then walk out to fly away. When they enter the hive entrance, most bees enter on the side nearest the cluster. After entering, they walk to the side and walk up the inside of the hive. It makes me think, what does the traffic flow look like inside the Langstroth hive all filled out with frames and foundation.

In light of this, maybe when we install packages, we should not center the queen cage, maybe we should be placing it nearer to one side to make their effective distance from the entrance a little shorter.

George

Through the Warre' Window, Part 2

Well, the hive mind changed. They had drawn out wax from one side over about 6 frames and it looked like they were going to draw wax in the lower box rather than fill the upper box. They shifted their tactics and filled the top box. Then they started working their way down in the bottom box. Unfortunately they have reached the bottom in the middle and the festooning bees are blocking my view. I may have to purchase another box soon with a window in it to add a new view. Maybe I could harvest some of the bees to add to other hives. (Anything to open up the view.)

I need to get working on my painting design for the hive. Plain white primer does not suit it well. And if I get another box to add, that will make me further behind in my maintenance.

Maybe I should open this hive up and look at the frames. I wonder if the bees have stuck the frames to the hive wall in the upper chamber. Also wonder if the comb in the upper box is attached to the top of the frames in the lower box. Note to self: For the next Warre' hive, get a window in both boxes. The window provides so much more.

George

Making lemonade with Mary's Bees. Jun 18, 2011.

I received a call from Mary whose bees were going crazy in the orchard. Lots of Bees in the air. Not forming into a swarm and leaving or settling, just having a gay old time in the warm afternoon. They eventually settled back into the hives. She said this was not normal activity. We agreed to take a look at them today to find out what was up. I also thought to bring some optional equipment that may be useful. I was hoping that it was just practice for a swarm where the queen refused to leave and I would find hives overflowing with bees, after all, this is always the desire. And by the way, Mary has 2 hives.

Slight mist today when we started and only an occasional bee would fly in or out of the hive. One hive had a tongue of bees sticking out the entrance, the other one did not. We opened the one with fewer bees first. The super of foundation was recently put on and not much activity in it so we set it off to the side. The deep box was full of bees. We found brood on 8 of 10 frames. We also found Multiple Queen cells on 4 of the frames. We also found a young queen walking around. This hive had obviously swarmed in the last week. With the amount of brood, in comparison to the current population, it was probably a pretty large swarm. The activity the day before may have been the young queen taking a flight. We isolated the queen on her frame, and isolated the frames with queen cells. Fortunately we had a light mist and the bees were staying put on the frames. Before deciding on what to do with this hive, we thought to look through the second hive.

The second hive was working on half of the frames in the super that was recently added. We checked a couple of the frames to verify no brood was in the super and then set the super off to the side. The deep box was very

full of bees. Also there were only 9 frames in it, but the bees had constructed their own frame-less comb structure in the area where the 10th frame should be. We started the inspection from the other side. This box obviously has more bees, as we had to nudge bees out of the way so we could see the frames. We found queen cells on several frames and removed them as we went. After looking at all 9 frames, we found lots of brood, young larva, maybe eggs, but no queen. With a slight mist and no sun, it is nearly impossible to see eggs. We smoked the frame-less frame with extra force so we could see. It was almost exclusively capped drone brood but had a queen cell on one spot. We were able to cut and pull it out in sections being careful not to crush the bees covering it. When the last section pulled loose, we found the queen walking around on it. She looked like the original queen and was looking good. We let her walk onto one of the other frames.

So now for the lemonade. We have one hive already swarmed with extra queen cells, and one hive exploding with population getting ready to swarm. (Pause a moment to think what you would do.) I always try to make use of queen cells if possible. The optional equipment I brought consisted of a Snelgrove board and a split deep box. A split deep box is a deep box with a divider down the middle to separate the compartment into two halves. Then each side can be used as a 4 frame nuc box. A Snelgrove board is like an inner cover, but the hole is screened over and there are key way entrances on all 4 sides, on both the upper and lower halves that can be opened or closed. The key way entrances can be opened or closed depending on what is needed. We will only open the top keyway on both sides to provide entrances for the split deep box. In each side of the split deep box, we put 3 frames making sure to mix frames from both hives. We made sure both sides have multiple queen cells. We also shook some extra bees into each side from the super that had lots of bees.

We then reassembled both hives, centering the frames and filling in the outside with foundation as to not let the bees create frame-less frames. As we had further reduced the first hive that had already swarmed, we moved a couple frames of bees from the stronger hive into the weaker one. As this had the young queen, we left two queen cells in the box. If this queen does not work out, the hive may have another option. We also left off the super as the bees have a ways to build back up before they need room for honey storage. This hive gets a feeder as it is mostly young bees, as the swarm took most of the foragers.

We left the super on the stronger hive as it still has most of its population. Also made sure the hive gets 10 frames in the brood chamber. The stronger hive keeps their super. Also, some of the bees we moved to the other hive will fly back from where we put them at their next opportunity. We put the Snelgrove board above the super of the stronger hive with the split deep containing two 3-frame nucs on top of that. Then we put a canvas liner and covered with a flat lid, stored the unused inner cover on top and added the metal cover to the top.

Now the goal is to leave them alone for 3 weeks and then check back. Ideally, the new queen will be laying up a storm in the first hive and it may need a super. The older queen in the strong hive will still be in the box and will not have swarmed. And in each side of the split deep, there will be young queens starting to lay. Maybe next month we will find out how it worked out.

We could have divided the split deep into 4 compartments and tried to make 4 new queens but I do not think Mary needs that many queens for only 2 hives.

George

George Imirie Essay Library

Master Beekeeper George Imirie (1933-2007) of Maryland published scores of helpful essays for beekeepers, including some specifically for the TBA. All files in Adobe Acrobat format.



<http://www.tnbeekeepers.org/learning.htm> (more essays here)

Essays

Colony Management During Nectar Flow

<http://www.tnbeekeepers.org/pubs/Colony%20Management%20during%20May%20Nectar%20Flow%20May%201999.pdf>

Bee Management

<http://www.tnbeekeepers.org/pubs/Bee%20Management%20October%202000.pdf>

Optimizing Honey Production

<http://www.tnbeekeepers.org/pubs/Optimizing%20Honey%20Production%20-%201999%20ABF%20Workshop.pdf>

Supering

<http://www.tnbeekeepers.org/pubs/Supering%20April%201999.pdf>

Finishing out Supers

<http://www.tnbeekeepers.org/pubs/Finishing%20out%20Supers%20June%201999.pdf>

Nectar Flow "Must Do" Items

<http://www.tnbeekeepers.org/pubs/Nectar%20Flow%20must%20do%20items.pdf>

Harvesting Honey

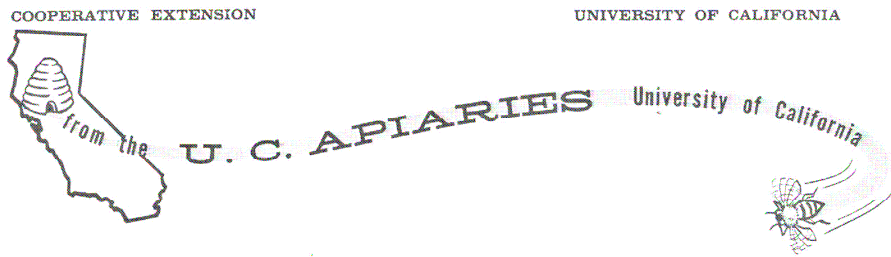
<http://www.tnbeekeepers.org/pubs/Harvesting%20Honey.pdf>

Ready For The Nectar Flow?

<http://www.tnbeekeepers.org/pubs/Ready%20for%20the%20Nectar%20Flow.pdf> ready for the Nectar Flow?

Proper Time and Technique of Harvesting Honey

<http://www.tnbeekeepers.org/pubs/Proper%20Time%20and%20Techniques%20of%20Harvesting.pdf>



Another Bee Gut Study

It seems like there is quite a burst of interest in the intestinal microbes residing in the honey bee digestive system and the possible contributions they make to pollen digestion both in the gut and in stored pollens in the combs (bee bread).

Until recently, studies of bacteria and fungi have been limited by the fact that many of them are "fastidious" and will grow only on very special, chemically unique "media." Also, much less attention has been paid to anaerobes, bacteria that cannot be grown in the presence of oxygen.

Now, however, many bacteria and fungi have been teased apart genetically and their DNA sequences recorded in giant data-bases. Even if the microbe turns out to be unknown, you can tell what its near relatives are (thank goodness for computer search-es!).

Additionally, when you study the sequences you can determine what enzymes the microbes should be able to produce and what sorts of food they can digest, etc.

Thus, a number of "bee labs" are becoming involved in studies of these microbes and what exposure to antibiotics and pesticides may be doing to them.

Now, interest has spread to labs that previously were not bee labs. Dr. DeRisi's lab in San Francisco is developing a micro-array chip for all known diseases, and other associates, of honey bees and other arthro-pods. Most recently, there has been a not-too-well-advertised plea from a lab at Yale University, in West Haven, CT, for beekeepers to collect about 20 live bees from the outside edge of the brood rearing cluster, into two provided tubes of (safe-to-handle) preservative for each of four colonies, and send them back to Yale for intestinal microbe identification. Again, this will be a genetic analysis, without growing microbes in the lab. Dr. Nancy Moran, lead investigator, wishes to receive samples from all over the U.S. and around the world, if possible. There is a questionnaire form on colony management to fill out for each colony, too.

If you wish to find out what microbes (of the eight groups they expect to find) are resident in your bees, go to the URL: <http://yale.edu/moran/bees.html> Click "You want us to send you a sample kit?" and you can request a kit containing eight tubes of preservative, stickers to link the sample tubes to the data sheets, the forms, packing, and return shipping address.

Colony Management Practices Survey

Do you wonder about some of your colony management practices? Are they helping or hurting your bees? How are other beekeepers dealing with problems and managing their bees? This information currently is being collected, sorted, grouped, analyzed and readied for beekeepers at one web site: <http://Beeinformed.org> Dennis vanEngels-dorp is spearheading an effort to collect all the good, the bad, and the ugly, including survival statistics, etc. and match them to colony maintenance practices. Before long, the Bee Informed Partnership site will allow beekeepers to compare their practices with those of thousands of other beekeepers, as well as look at the success of those other practices. Beekeeping operations, from one in the backyard to tens of thousands used commercially, should be included in the data-base. This compilation will be valuable only if there are very large numbers of responses to compare. If you can find a few minutes to complete the forms, electronically, at the bottom of the page at <http://Beeinformed.org> you will be contributing to a huge data set and will help others, as well as yourself, to determine what are apt to be the best practices for managing your colonies.

Thank you for taking the time to share your management practices and results.

WASHINGTON STATE BEEKEEPERS ASSOCIATION

Keep the Bee in Business

<http://www.wasba.org/>

Honey Bee Field Day

June 24-25, 2011 WSU Campus - Ensminger Pavilion and Apiaries

Registration form

Word: <http://www.wasba.org/2011%20Field%20Day%20Reg%20Form%5B1%5D.doc>

PDF: <http://www.wasba.org/2011%20Field%20Day%20Reg%20Form1.pdf>

Field Day Topics PDF:

[http://www.wasba.org/2011%20WSU%20Honey%20Bee%20Field%20Day%20Topics\(final%20from%20Steve\).pdf](http://www.wasba.org/2011%20WSU%20Honey%20Bee%20Field%20Day%20Topics(final%20from%20Steve).pdf)

Latest update on request for HopGuard for Varroa control

Please take a look at the letter to EPA here:

<http://www.wasba.org/Request%20for%20use%20of%20HopGuard%20on%20honey%20bee%20colonies%20to%20control%20varroa%20mites%20082410.pdf>

PNW Mortality Survey

Oregon State University PNW Mortality Survey. If you keep bees in the Pacific Northwest, please take a few minutes to complete this short survey. Dr. Mike Burgett's mailing address is at the bottom of the document. Thank you!

<http://www.wasba.org/Colony%20Mortality%20Questionnaire%20-%20May%2009.pdf>

Honey Bee Diagnostics Lab

Washington State beekeepers now have a Honey bee diagnostic lab right at home. For more information:

http://apis.wsu.edu/diagnostic_center.html

Use these forms to submit samples for testing:

Colony Health data sheet (pdf)

http://apis.wsu.edu/2009%20updates/documents/ColonyHealthDataSheet_000.pdf

Field Sampling Protocol (pdf)

http://apis.wsu.edu/2009%20updates/documents/FieldSamplingProtocol_000.pdf

Bee News Links

Picky Pollinators: Native Bees are Selective about Where They Live and Feed

http://www.usgs.gov/newsroom/article.asp?ID=2833&from=rss_home

How honeybees control their flight speed to avoid obstacles

<http://www2.cnrs.fr/en/1861.htm>

How Bees Learn Which Odors to Follow

http://www.fu-berlin.de/en/presse/informationen/fup/2011/fup_11_055/index.html



<http://www.sciencedaily.com/>

Honey Can Reverse Antibiotic Resistance, Study Suggests (Apr. 13, 2011) — Manuka honey could be an efficient way to clear chronically infected wounds and could even help reverse bacterial resistance to antibiotics, according to new ... > <http://www.sciencedaily.com/releases/2011/04/110412201713.htm>

How Manuka Honey Helps Fight Infection (Sep. 10, 2009) — Manuka honey may kill bacteria by destroying key bacterial proteins. Researchers investigated the mechanisms of manuka honey action and found that its anti-bacterial properties were not due solely to ... > <http://www.sciencedaily.com/releases/2009/09/090907013759.htm>

Patients Should Ask Surgeons About Using Honey To Heal Wounds (Oct. 19, 2007) — Honey is enjoying a resurgence as a wound-healing solution amid rising concerns about antibiotic resistance and a renewed interest in natural healing. Researchers started to document its success in ... > <http://www.sciencedaily.com/releases/2007/10/071017102057.htm>

Honey Helps To Heal Wounds, Review Suggests (Oct. 9, 2008) — Honey may reduce healing times in patients suffering mild to moderate burn wounds. A systematic review concluded that honey might be useful as an alternative to traditional wound dressings in ... > <http://www.sciencedaily.com/releases/2008/10/081007192524.htm>

Honey as an Antibiotic: Scientists Identify a Secret Ingredient in Honey That Kills Bacteria (July 12, 2010) — Sweet news for those looking for new antibiotics: new research explains for the first time how honey kills bacteria. Specifically, the research shows that bees make a protein that they add to the ... > <http://www.sciencedaily.com/releases/2008/10/081007192524.htm>

BEEKEEPERS IN THE SOUTH

On a recent trip to Georgia we had the good fortune to attend the monthly meeting of the Coastal Empire Beekeepers Association in Savannah. Held at the home of Fred and Liz Grier, the meeting drew a crowd of around 45 members and guests.

The main item on the agenda was a presentation by two representatives of the Georgia Department of Agriculture who explained the State's position on and requirements of hobbyist, sideline and small commercial beekeepers. Without going into detail on the requirements, it seems as though Georgia's regulations on the sales of local honey are more restrictive than Washington State's. During the meeting the members asked some excellent and interesting questions and the DOA representatives did a fine job of answering and even included handouts and a Power Point presentation.

Also present at the meeting were around a thousand honeybees in a lovely two-frame observation hive.

Prior to the meeting we had the opportunity to tour Fred's 9+ acre place (via club car) and take a look at his 30 hives – nine of which were "rescues" from the local US Post Office. Seems a honeybee distributor shipped bees to the local post office and there were an enormous number of "hitchhikers" which were taken in by members of their Association. Most of Fred's hives are Langstroth but he also has a couple nucs and beautiful new top bar hive. Fred's bees produced a delightful, light honey and a robust dark honey – both of which were available for sampling at the June 21st WSBA meeting along with Basil's thick wildflower comb honey and some Georgia wildflower comb honey (another good reason to come to the monthly meetings!)

After quizzing some of the members we were able to determine that their honey production is mainly wildflower with a strong influence of Chinese Tallow and, in some areas, Tupelo. This year they have had very little rainfall and, as we all know, lack of rain = lack of nectar = lack of honey so we wished some of our rain on them (to be administered AFTER we left, of course.) We left Savannah on Wednesday afternoon and, that midnight, the clouds rolled in and gave them a much needed drenching... (glad we could help!)

Members of the CEBA were extremely gracious and showed us the famous Southern hospitality that we've come to know and love from Savannah. We're already looking forward to our next trip and we hope it coincides with another CEBA meeting!

Advanced Beekeeping in the Pacific Northwest 2011

Join us for our Advanced Beekeeping Course Saturday July 9th, 2011 8:30am-4:30pm here at our West Sound Beekeeping Association located at Stedman Bee Supplies. This course is for seasoned beekeepers and for people that have already taken the Basics course and need a refresher course.



Some beekeepers have kept bees for years, but because they keep repeating the same mistakes they are not gaining years of beekeeping experience. Instead they are stuck in being a first year beekeeper year after year. Take the next step, and leap into becoming a better beekeeper! We'll take a more in depth look at swarm prevention, splits, overwintering hives, pests & disease diagnosis, treatment and prevention, increase honey yields and tricks of the trade.

There will be hands on demonstrations in the afternoon so all beekeepers must bring a hat and veil or suit. For example you will have the opportunity to participate in activities such as making splits, sugar rolls, sugar dusting and mite counts, hygienic testing, etc.



Beekeeping in the Pacific Northwest 2011

Saturday July 9, 2011
Silverdale, Washington

Learn about:

ADVANCED BEEKEEPING CLASS

- European Honey Bee Biology
- Types of Flowers Used
- Equipment Use Review
- Workings of the Hive
- Swarm prevention
- Splits
- Varroa Mite, Tracheal Mite & Nosema
- Seasonal Management
- Pulling the Surplus Honey
- The Extraction Process
- Honey Marketing

Agenda

Saturday, July 9, 2011

8:00 am – 8:30 am
8:30 am - 4:30 pm

Registration / Check in
Class (lunch provided) *see topics above*

Instructors:

Paul Lundy, President, Washington State Beekeeping Association

George Purkett, West Sound Beekeepers Association

David Mackovjak, Queen Rearing Chairman, West Sound Beekeepers Association

Guest Speakers from the West Sound Beekeepers Association and from other areas of the beekeeping industry.

Registration: We expect this class to fill early and early registration is recommended. Please check our website for availability -- <http://www.westsoundbees.org> . Questions or concerns can be addressed to David Mackovjak, queen_rearing@westsoundbees.org or 360-340-0381

Download Registration form (pdf)

\$50.00, registrations on or before July 1, 2011

\$70.00, registrations after July 1, 2011

(Fee includes Saturday lunch, refreshments, and course materials)

Refunds Policy: \$25.00 will be refunded to participants who cancel in writing prior to July 6, 2011.

Cancellations: Send cancellations to Kim Redmond, WSBA Treas., P.O. Box 536 Chimacum, WA 98325, or by email – biz@kimredmond.com

Location: West Sound Beekeeping Association located at Stedman's Bee Supplies, 3763 NW Anderson Hill Rd Silverdale, WA 98383

Stedman's Bee Supplies:



<http://tinyurl.com/3mwepna>

Silverdale, WA:



<http://tinyurl.com/3p84dvd>

Nearby Lodging

- **Silverdale Beach Hotel**, Silverdale, WA (360) 698-1000 <http://www.silverdalebeachhotel.com/>
- Oxford Suites Hotel, Silverdale, WA (360) 698-9550 <http://www.oxfordsuitessilverdale.com/>

Advanced Beekeeping in the Pacific Northwest 2011

Registration Form

Saturday July 9, 2011
Silverdale, Washington

Please use a separate form for each registration.

Method of Payment: check or money order

Registration Fee:

Enclosed is a check or money order.

\$50.00 if postmarked on or before July 1, 2011

\$70.00 postmarked after July 1, 2011

Name _____

Address _____

City _____

State _____ Zip _____

Phone _____

E-mail _____

Complete the registration form above, include payment and mail to:

Kim Redmond
WSBA Treasurer
P.O. Box 536
Chimacum, WA 98325